THE RELATIVE CONTRIBUTIONS OF CYSTATHIONINE \( \beta \)-SYNTHASE AND \( \gamma \)-CYSTATHIONASE TO \( \text{H}_2\text{S} \) BIOGENESIS VIA ALTERNATIVE TRANSULFURATION REACTIONS \(^1\)

Sangita Singh, Dominique Padovani, Rachel A. Leslie, Taurai Chiku and Ruma Banerjee

\(^1\)Department of Biological Chemistry, University of Michigan Medical Center, Ann Arbor, MI 48109

Running Title: Biogenesis of \( \text{H}_2\text{S} \) from CBS and CSE

\(^*\)Address correspondence to: Ruma Banerjee, 3320B MSRB III, 1150 W. Medical Center Dr., University of Michigan, Ann Arbor, MI 48109-5606, Tel: (734) 615-5238; E-mail: rbanerje@umich.edu
In mammals, the two enzymes in the transsulfuration pathway, cystathionine β-synthase (CBS) and cystathionine γ-lyase (CSE), are believed to be chiefly responsible for \( \text{H}_2\text{S} \) biogenesis. In this study, we report a detailed kinetic analysis of the human and yeast CBS-catalyzed reactions that result in \( \text{H}_2\text{S} \) generation. CBS from both organisms show a marked preference for \( \text{H}_2\text{S} \) generation by \( \beta \)-replacement of cysteine by homocysteine. The alternative \( \text{H}_2\text{S} \)-generating reactions, i.e. \( \beta \)-elimination of cysteine to generate serine or condensation of two moles of cysteine to generate lanthionine are quantitatively less significant. The kinetic data were employed to simulate the turnover numbers of the various CBS-catalyzed reactions at physiologically relevant substrate concentrations. At equimolar concentrations of CBS and CSE, the simulations predict that \( \text{H}_2\text{S} \) production by CBS would account for ~25-70% of the total \( \text{H}_2\text{S} \) generated via the transsulfuration pathway depending on the extent of allosteric activation of CBS by S-adenosylmethionine. The relative contribution of CBS to \( \text{H}_2\text{S} \) genesis is expected to decrease under hyperhomocysteinemic conditions. CBS is predicted to be virtually the sole source of lanthionine and CSE, but not CBS, efficiently cleaves lanthionine. The insensitivity of the CBS-catalyzed \( \text{H}_2\text{S} \)-generating reactions to the grade of hyperhomocysteinemia is in stark contrast to the responsiveness of CSE and suggests a previously unrecognized role for CSE in intracellular homocysteine management. Finally, our studies reveal that the profligacy of the transsulfuration pathway results not only in a multiplicity of \( \text{H}_2\text{S} \)-yielding reactions but also yields novel thioether metabolites, thus increasing the complexity of the sulfur metabolome.

Hydrogen sulfide (\( \text{H}_2\text{S} \)) elicits an array of physiological effects including modulation of blood pressure and reduction of ischemia reperfusion injury (1,2). Other novel effects of \( \text{H}_2\text{S} \) include induction of a state of suspended animation in mouse by decreasing oxygen consumption and drastically reducing the metabolic rate (3) and synchronizing ultradian metabolic oscillation in yeast (4). Under conditions of metabolic cycling in yeast, \( \text{H}_2\text{S} \) production is catalyzed by sulfite reductase in the sulfur assimilation pathway (4). Inhibition of sulfite reductase reduces \( \text{H}_2\text{S} \) production and in turn, perturbs metabolic oscillations. \( \text{H}_2\text{S} \) is a specific and potent inhibitor of cytochrome c oxidase in the electron transport chain (3).

Although concentrations of \( \text{H}_2\text{S} \) have been reported to range from 50-160 µM in brain (5-7) and 30-50 µM in the peripheral system (8), these appear to be grossly overestimated (9). Significantly lower \( \text{H}_2\text{S} \) concentrations of 17 and 14 nM in liver and brain respectively, have been reported recently (9). The very significant discrepancy between these and the previous estimates of \( \text{H}_2\text{S} \) levels presumably derives from the earlier use of acidic conditions that led to the release of acid-labile sulfur from iron-sulfur centers.

In mammals, the primary catalysts for \( \text{H}_2\text{S} \) generation are reported to be the two pyridoxal phosphate (PLP)-dependent enzymes involved in the transsulfuration pathway, cystathionine β-synthase (CBS) and cystathionine γ-lyase (CSE) (10,11). The transsulfuration pathway operates in the reverse direction in mammals serving to convert homocysteine to cysteine (Fig. 1) while in yeast and bacteria the pathway is involved in sulfur assimilation from sulfate to cysteine. CBS is widely assumed to be the major contributor to \( \text{H}_2\text{S} \) production in the brain due to its relatively high expression in this organ (10). However, a recent study reported that 3-mercaptoppyruvate sulfurtransferase together with cysteine aminotransferase might also generate \( \text{H}_2\text{S} \) in brain (12). The relative contributions of these enzymes and of CSE, which is also present in brain (13,14), to \( \text{H}_2\text{S} \) production remain to be assessed. Genetic disruption of CSE in mouse leads to cardiac deficits including pronounced hypertension and reduced endothelium-dependent vasorelaxation, consistent with a major role for CSE in the peripheral system (1). However, brain \( \text{H}_2\text{S} \) levels are reportedly unchanged in CSE\(^{-/-}\) mice.

Despite the growing recognition of the varied physiological effects of \( \text{H}_2\text{S} \), our understanding of its regulation and mechanism of its biosynthesis is poor. We have recently reported on the complex kinetics of \( \text{H}_2\text{S} \) generation by human CSE (15).
The profligacy of the human enzyme affords \( \text{H}_2\text{S} \) generation by a multiplicity of routes involving cysteine and/or homocysteine as substrates. Kinetic simulations predict an increasingly important contribution of homocysteine to \( \text{H}_2\text{S} \) generation with increasing grade of hyperhomocysteinemia, a risk factor for cardiovascular and neurodegenerative diseases (16-18). In addition to \( \text{H}_2\text{S} \), a variety of products are generated in these reactions including two novel sulfur metabolites, lanthionine and homolanthionine, which represent the condensation products between two moles of cysteine and homocysteine respectively. Although the steady-state kinetic parameters for \( \text{H}_2\text{S} \) generation from cysteine and homocysteine have been reported for human CBS (hCBS) (19), a comparable detailed kinetic analysis of \( \text{H}_2\text{S} \) generation by CBS by multiple pathways and their sensitivity to the grade of hyperhomocysteinemia is not known. Furthermore, the relative contributions of CBS and CSE to \( \text{H}_2\text{S} \) and lanthionine generation at physiologically relevant concentrations of substrate are not known.

Human CBS is a unique heme containing, PLP-dependent enzyme (20) that catalyzes the \( \beta \)-replacement of serine by homocysteine to produce cystathionine. The latter is further metabolized by CSE in an \( \alpha,\gamma \)-elimination reaction to produce cysteine. Although yeast and human CBS are highly homologous and catalyze the same chemical reaction with similar kinetic parameters, the yeast enzyme lacks heme and is not allosterically regulated by S-adenosylmethionine (AdoMet) (21).

In the present study, we have elucidated the kinetics of \( \text{H}_2\text{S} \) biogenesis by yeast and human CBS and used simulations to estimate the relative contributions of CBS and CSE to \( \text{H}_2\text{S} \) production at physiologically relevant concentrations of substrate. We find that although CBS and CSE share a common feature, i.e. catalytic promiscuity. However, in contrast to CSE which is proficient at catalyzing reactions at the \( \beta \) and \( \gamma \)-carbons of substrates (15), CBS activity is confined to chemical transformations at the \( \beta \)-position. Our studies provide new insights into the existence of alternative transsulfuration reactions that can be a source of diverse sulfur metabolites, viz. \( \text{H}_2\text{S} \), lanthionine and homolanthionine increasing the diversity of the sulfur metabolome.

**Experimental Procedures**

**Chemicals**—L-Cysteine, D,L-homocysteine, D,L-lanthionine, L-serine, kanamycin and ampicillin were purchased from Sigma.

**Expression and Purification of hCBS, yCBS and human CSE**—Recombinant full-length hCBS and yeast CBS (yCBS) were purified as described previously (22,23). The expression plasmids encoding hCBS and yCBS were provided by Drs. Warren Kruger (Fox Chase Cancer Center) and Edith Miles (NIH, Bethesda) respectively. The expression plasmid for human CSE was provided by Dr. Markus Wahl (Max Planck Institute, Mantinsried, Germany). Recombinant human CSE was purified as described previously (15).

**Enzyme Activity Assays**—The following assays were used to determine CBS activity. In all assays, the concentration of the variable substrate ranged from 0.2×\( K_M \) to 5-20×\( K_M \). One unit of activity is defined as one micromole of product formed min\(^{-1}\) mg\(^{-1}\) of protein. For assays with hCBS, the allosteric activator, AdoMet (360 \( \mu \)M), was added to the reaction mixture.

**Radiolabeled assay**—The activity of hCBS and yCBS were measured in the radioactive assay using \([\text{\textsuperscript{14}}\text{C}]\)-serine as described previously (24).

**Detection of \( \text{H}_2\text{S}-\text{H}_2\text{S} \) generation from cysteine or from cysteine plus homocysteine** was measured in a spectrophotometric assay as described previously (15). The reaction mixture containing 100 mM Hepes buffer pH 7.4, 0.4 mM lead acetate, and varying concentrations of substrate (cysteine ± homocysteine) was preincubated at 37°C for 2 min, and the reaction was initiated by addition of yCBS. For hCBS, the enzyme was preincubated with 360 \( \mu \)M AdoMet (final concentration) at 37°C for 2 min, and the reaction was started by addition of substrate. Lead acetate (0.4 mM) did not inhibit CBS activity as determined in the radioactive assay (data not shown). A molar extinction coefficient of 5,500 M\(^{-1}\) cm\(^{-1}\) was used for lead sulfide as described previously (15).
HPLC analysis of serine and lanthionine-HPLC was used to estimate the rate of serine and lanthionine formation at varying concentrations of cysteine. The concentration of serine or lanthionine was determined following o-phthalaldehyde derivatization as described previously (15). Under these conditions, lanthionine and serine eluted with retention times of 3.2 min and 4.2 min, respectively. This HPLC method was also used to determine whether homoserine (5.26 min retention time) was produced by the CBS-catalyzed γ-elimination of homocysteine.

Detection of α-ketoacid products-CBS catalyzed generation of H₂S from cysteine or homocysteine could potentially generate the α-ketoacids, pyruvate and α-ketobutyrate. The colorimetric assay for detection of α-ketoacids was employed as described previously (15).

Detection of cysteine-The DTNB assay was used to measure the CBS- and CSE-catalyzed production of cysteine from lanthionine as described previously (15).

Estimation of Dissociation Constant for Cysteine and Serine-Fluorescence titrations were performed at 25°C on a Shimadzu RF-5301PC spectrofluorimeter. Human CBS (0.2 mg/ml) in 100 mM Hepes buffer, pH 7.4 containing 360 μM AdoMet, was incubated with varying concentrations of L-cysteine or L-serine. After 3 min incubation, spectra were acquired with an excitation wavelength of 400 nm. Data were analyzed by plotting ΔF₄₉₅nm as a function of cysteine or serine concentration and the Kₐ was determined by fitting the data with the Michaelis-Menten equation.

Mass Spectrometric (MS) Analysis of Reaction Products-Mass spectrometric analysis of lanthionine was performed described previously (15).

Kinetic Analysis-H₂S-generating reactions catalyzed by CBS (2-4) are either unimolecular (2) or bimolecular (3 and 4). Thus Kₘ₁ and Vₘₐₓ₁ refer to the kinetic parameters associated with the unimolecular reaction whereas Kₘ₂ and Vₘₐₓ₂ refer to substrate binding at the second site and the reaction velocity of the bimolecular reaction, respectively.

Serine and lanthionine generation-The Kₘ and Vₘₐₓ values for CBS-catalyzed serine (reaction 2) or lanthionine (reaction 3) production from cysteine were determined using the HPLC assay as described above and data were fitted using equations 1 and 2-4 respectively.

\[
V_{\text{Serine}} = \frac{V_{\text{max}} [\text{Cys}]}{K_{M1(\text{Cys})} + [\text{Cys}]} \left(1 + \frac{[\text{Cys}]}{K_i}\right)
\]

\[
V_{\text{Lanthionine}} = \frac{V_{\text{max}} 2 [\text{Cys}]^2 [\text{Cys}]^n}{[\text{Cys}]^2 + K_{M1(\text{Cys})} [\text{Cys}]^n + [\text{Cys}]K_{M2}^n} [\text{Cys}]^n
\]

As binding of cysteine to the second binding site leads to depletion of substrate for the unimolecular reaction, a substrate inhibition term [Cys] × (1+[Cys]/Kᵢ) was included in equation 1. In principle, the bimolecular reaction catalyzed by CBS can follow a binary (ping-pong) or sequential/ternary (random or ordered) mechanism. Hence, the experimental data for lanthionine production from cysteine (reaction 3) were fitted using equations 2, 3 or 4 that describe a ping-pong, random-sequential, and ordered-sequential mechanism respectively. As cooperativity was clearly observed with γCBS for binding of substrate to site 2, a Hill coefficient (n) was included in these equations.

\[
V_{\text{Lanthionine}} = \frac{V_{\text{max}} 2 \left(1 + \frac{K_{M1(\text{Cys})} [\text{Cys}]}{[\text{Cys}]^2} + \frac{K_{M2(\text{Cys})}^n [\text{Cys}]^n}{[\text{Cys}]^2 + [\text{Cys}]K_{M2}^n} \right)}{1 + \frac{K_{M1(\text{Cys})} [\text{Cys}]}{[\text{Cys}]^2} + \frac{K_{M2(\text{Cys})}^n [\text{Cys}]^n}{[\text{Cys}]^2 + [\text{Cys}]K_{M2}^n} + \frac{K_{d1(\text{Cys})}}{[\text{Cys}]^2}}
\]

H₂S production from cysteine-H₂S production from cysteine by CBS is the sum of a unimolecular (reaction 2) and a bimolecular (reaction 3) reaction. The latter can proceed via a ping-pong or sequential (ordered or random) mechanism as discussed above. Hence, the experimental data for H₂S production from cysteine (i.e. reactions 2-3) monitored by the continuous lead acetate assay described above were fitted using equation 5 with V_{serine} described by equation 6 and V_{lanthionine} described by equations...
2, 3 or 4 to account for the alternative mechanisms.

\[ v_{H_2S} = v_{\text{Serine}} + v_{\text{Lanthionine}} \]  

\[ v_{\text{Serine}} = \frac{V_{\text{max}}[\text{Cys}]}{K_{M1(\text{Cys})} + [\text{Cys}]} \times \frac{1}{1 + \frac{[\text{Cys}]}{K_{M2(\text{Cys})}}} \]  

Binding of cysteine to the second binding site (reaction 3) will affect the \( V_{\text{max}} \) values for \( H_2S \) formation in the unimolecular reaction 2. Hence, an inhibition term, where \( K_i \) is assumed to be equal to \( K_{m2} \), was introduced in equation 6. This assumption is consistent with the high \( K_i(\text{Cys}) \) (11 mM) reported for cysteine with respect to homocysteine for the CBS-catalyzed reaction by rat CBS (25).

**H\(_2\)S production from cysteine and homocysteine-CBS** does not catalyze detectable \( H_2S \) formation from homocysteine alone as monitored in the lead acetate assay. However, in the presence of cysteine, CBS catalyzes a condensation reaction between these two amino acids (4). Hence, the kinetic parameters for reaction 4 were determined by fitting the experimental data for \( H_2S \) formation obtained in the presence of 20 mM cysteine for hCBS or 15 mM cysteine for yCBS and varying the concentration of homocysteine. In this set of experiments, the observed rate of \( H_2S \) production represents the sum of reactions 2, 3 and 4, as described by equation 7. The values for \( v_2 \to v_4 \)

\[ v_{H_2S} = v_2 + v_3 + v_4 \]  

corresponding to the reaction velocities for 2, 3 and 4 respectively were obtained using equations 8-10 for the ping-pong mechanism.

\[ v_2 = \frac{V_{\text{max}}[\text{Cys}]}{K_{M1(\text{Cys})} + [\text{Cys}]} \times \frac{1}{1 + \frac{[\text{Cys}]}{K_{M2(\text{Cys})}}} \times \frac{1}{1 + \frac{[\text{Hcy}]}{K_{M2(\text{Hcy})}}} \]  

\[ v_3 = \frac{V_{\text{max}}[\text{Cys}][\text{Cys}]}{[\text{Cys}][\text{Cys}]} \times \frac{1 + \frac{[\text{Hcy}]}{K_{M2(\text{Hcy})}}}{K_{M1(\text{Cys})}[\text{Cys}]} \]  

\[ v_4 = \frac{V_{\text{max}}[\text{Cys}][\text{Hcy}]}{[\text{Cys}][\text{Hcy}]} \times \frac{1 + \frac{[\text{Cys}]}{K_{M2(\text{Cys})}}}{K_{M1(\text{Hcy})}[\text{Hcy}]} \]

In these equations, \( n \) and \( k \) represent the Hill coefficient for the binding of the second substrate in reactions 3 and 4 respectively. The kinetic parameters obtained for reactions 2 and 3 as described above were constrained to obtain the kinetic parameters for reaction 4. A term for non-competitive inhibition was introduced in equation 8, assuming that \( K_i=K_{m2} \), since the simultaneous presence of both substrates will lead to inhibition of the unimolecular reaction. Also, a competitive inhibition term for the binding of the second substrate was included since cysteine and homocysteine will compete for binding at the second binding site to catalyze different bimolecular reactions. We assumed that \( K_i=K_{m2} \).

**Results**

**Initial Analysis of \( H_2S \) Production by hCBS and yCBS**—The specific activity of recombinant hCBS used in these studies was 5.1±0.3 µmol cystathionine formed min\(^{-1}\)mg\(^{-1}\) of protein at 37°C and pH 7.4 when homocysteine and serine were employed as substrates and is comparable to the value published previously (22). The specific activity of yCBS was 24±2 µmol cystathionine formed min\(^{-1}\)mg\(^{-1}\) and is similar to the value reported previously (23).

The kinetics of \( H_2S \) production by yeast or human CBS were characterized using the continuous spectrophotometric assay. Under \( V_{\text{max}} \) conditions, the condensation of cysteine and homocysteine represents the preferred pathway for \( H_2S \) production, as reported previously for hCBS (19), and specific activities of 60±2.4 U/mg for yCBS and 18.7±2.6 U/mg for hCBS were obtained (Table 1). These specific activities are ~7-fold and 23-fold higher than those observed for the bimolecular reaction with cysteine as substrate for yeast (9.0±1.3 U/mg) and human (0.82±0.08) CBS respectively.

**Product Analysis of \( H_2S \) Producing Reactions**—To distinguish between the multiple routes for \( H_2S \) generation (Fig. 2, reactions 2-6), reaction products were analyzed by UV-visible spectroscopy, mass spectrometry and HPLC. When cysteine was used as a substrate, serine and lanthionine formation were detected by HPLC and
the presence of the latter was confirmed by mass spectrometry. Two daughter ion peaks obtained by MS/MS analysis of the parent ion peak (m/z = 209) were assigned: m/z = 120 (corresponding to SCH₂CH(NH₂)COOH) and m/z = 192 (corresponding to loss of NH₃ from lantionine) (data not shown). Detection of serine and lanthionine are consistent with β-elimination (2) and β-replacement (3) reactions with cysteine as a substrate. In principle, serine formed in the unimolecular reaction (2) could be further converted to pyruvate and NH₃ as observed with CSE (15). However, the α-ketoacid assay failed to reveal the break down product of serine, i.e. pyruvate. This is consistent with the earlier report that neither chicken nor rat liver CBS catalyzes α,β-elimination reactions (26). Unlike CSE, CBS does not appear to use homocysteine by itself for H₂S production since none of the expected reaction products, i.e. H₂S, homoserine or α-ketobutyrate were detected. This is also consistent with fluorescence titration experiments, which failed to reveal evidence for binding of homocysteine to hCBS (not shown).

**Kinetics of H₂S Generation by yCBS**-The dependence of the rate of H₂S formation on cysteine concentration is markedly biphasic (Fig. 3A), which facilitated deconvolution of the kinetic parameters Vₘₐₓ and Kᵢₑₐₚₚ associated with the two phases representing reactions 2 and 3 respectively (Table 1). The kinetic data obtained for the bimolecular reaction (3) were fitted for alternative mechanisms (i.e. binary versus ternary) (Table S1). Based on the quality of the fits, a distinction between a ping-pong and random-sequential mechanism could not be made. In contrast, an ordered-sequential mechanism provided a poor fit to the data. The active site of CBS can bind two amino acids, serine and homocysteine, to catalyze the canonical β-replacement reaction (1). These sites are referred to as site 1 (i.e. site at which the external aldimine with PLP is formed) and site 2 respectively. Yeast CBS exhibits an ~10-fold lower Kᵢₑₐₚₚ for cysteine at site 1 (3.6±1.7 mM) compared to site 2 (33.0±3.7 mM) and positive cooperativity for binding of the second mole of cysteine is seen (n = 2.4±0.5). Under Vₘₐₓ conditions, the rate of the bimolecular reaction (3) is ~2-fold higher than for the unimolecular reaction (2) (Table 1).

The kinetics of reaction 4, i.e. the condensation of cysteine and homocysteine to form cystathionine and H₂S, is shown in Fig. 3B. The kinetic values for reaction 4 were determined by deconvolution of the three phases contributing to the net rate of H₂S formation, using as input values, the kinetic parameters determined in the presence of cysteine only as a substrate (Table 1). The kinetic data were fitted with alternative mechanisms and a reasonable fit was only obtained for the ping-pong mechanism with cysteine binding first (Table S2). The Kᵦₗₑₐₚₚ value for site 2 obtained from this analysis is ~26-fold lower than the Kᵦₗₑₐₚₚ value obtained for the condensation of serine and homocysteine (1) (Table 1). The reason for this difference is not known.

The turnover number under Vₘₐₓ conditions (i.e. kₑₐₚₚ) for the condensation of cysteine and homocysteine (4) is ~2-fold higher than for the canonical condensation reaction of serine and homocysteine (1). Reaction 4 represents a significantly more efficient route for H₂S generation than the competing reactions exhibiting an ~16- and 6.7-fold higher kₑₐₚₚ than reactions 2 and 3 respectively.

**Kinetics of H₂S Generation by hCBS**-The dependence of the rate of H₂S formation on cysteine concentration is not as markedly biphasic as observed with yCBS (Fig. 4A). Thus, the kinetic parameters for serine (reaction 2) and lanthionine (reaction 3) formation were experimentally determined at increasing concentrations of cysteine (Fig. 4B). The resulting Kᵢₑₐₚₚ value obtained for reaction 2 (Table 1) was then employed as an input parameter to fit the experimental data for H₂S production (Fig. 4A). The Kᵢₑₐₚₚ for cysteine binding to site 1 (6.8±1.7 mM) is ~4-fold lower than for site 2 (27.3±3.7 mM) for reaction 3. The kinetic data for H₂S production observed with cysteine and homocysteine as substrates are shown in Fig. 4C. As with yCBS, the kinetic data observed for the bimolecular reactions, i.e. reactions 3 and 4, were fitted with alternative mechanisms (Tables S2-S3). The outcomes of the fits were similar to those observed with yCBS and ruled out an ordered-sequential mechanism. We note that all the hCBS-catalyzed reactions were measured in the presence of the allosteric activator, AdoMet. As reported
previously (19), H₂S generation by hCBS exhibits a similar sensitivity to activation by AdoMet as does the canonical condensation reaction of serine and homocysteine (1) (data not shown).

Under Vₘₐₓ conditions, the turnover number for reaction 4 is 3.7-fold higher than for the condensation reaction of serine and homocysteine (1). As with yCBS, reaction 4 is a much more significant contributor to H₂S production being ~41- and ~23-fold more efficient than reactions 2 and 3 respectively. The observed increase in the specific activity of hCBS with cysteine versus cysteine and homocysteine as substrates is similar to the 30-fold difference reported by Braunstein and colleagues (27). The turnover number of H₂S generation from reaction 4 is ~2.8-fold higher for yeast versus human CBS (Table 1). The relative productivity of the three H₂S generating reactions follow the order: 4>3>2 for both yeast and human enzymes.

The Relative Contributions of the hCBS-catalyzed Reactions to H₂S Generation-The primary role of CBS in the transsulfuration pathway as widely accepted is to catalyze the condensation of serine and homocysteine to generate cystathionine (Fig. 1). Therefore, the higher turnover number for cystathionine production via reaction 4 (from cysteine + homocysteine) versus 1 (from serine + homocysteine) is surprising, and suggests the existence of alternative transsulfuration routes, whose operation must be regulated by mechanisms that await elucidation. Since the intracellular substrate concentrations are low compared to their respective Kₛₛ (serine, cysteine and homocysteine concentrations are assumed to be 560 μM, 100 μM and 10 μM, respectively), the contribution of the various CBS-catalyzed reactions to the overall pool of H₂S and cystathionine will be determined by substrate availability. Thus, to determine the partitioning between the three H₂S-producing reactions (2-4) and the two cystathionine-producing reactions (1 and 4), simulations were performed at the relevant physiological substrates concentrations (Table 2) using the kinetic parameters reported in Table 1. Furthermore, the relative contributions of each of the reactions to total H₂S or cystathionine production were estimated at three concentrations of homocysteine mimicking normal (10 μM) versus moderate (40 μM) and severe (200 μM) hyperhomocysteinemia (Tables 2 and 3). In this calculation, a competitive inhibition term (1+ [Ser]/Kₛ) was introduced in equations 8-10, since serine with a Kₛ of 56±6 μM will compete with cysteine to bind to the PLP site, i.e. site 1. Similarly, a competitive inhibition term (1+ Cys/Kₛ) was used for reaction 1 since cysteine exhibits a Kₛ of 400±27 μM. We note that both Kₛ values were determined at 25°C as described under Experimental Procedures and are likely to be higher at 37°C where the enzymatic activity was determined. Unfortunately, attempts to obtain the Kₛ values at 37°C resulted in poor data quality. Based on these values, the turnover number (v/[E]) for cystathionine production from serine and homocysteine (1) is predicted to be ~1.3-fold lower at 10 μM homocysteine but ~10.5-fold higher at 200 μM homocysteine than from cysteine and homocysteine (4). Thus, at low homocysteine concentrations, reactions 1 and 4 are predicted to contribute to cystathionine production at a ratio of ~2.3.

Next, we assessed the relative contributions of the hCBS-catalyzed H₂S-generating reactions at three concentrations of homocysteine (Tables 2 and 3). Our simulations predict that the condensation of cysteine and homocysteine (4) is the major contributor, accounting for ~96% of H₂S production, which is largely invariant over a 20-fold range of homocysteine concentration. The combined contribution of reactions 2 and 3 is 0% of negligible importance (Table 3). Importantly, the competition between serine and cysteine rather than homocysteine for site 1 on CBS, makes H₂S production by CBS relatively insensitive to homocysteine concentration, increasing only 8% as the concentration of homocysteine is increased from 10 to 200 μM (Table 2).

Predicted relative contributions of CBS versus CSE to H₂S generation-In contrast to CBS, the CSE-catalyzed generation of H₂S is responsive to the grade of hyperhomocysteinemia (15). Comparison of the turnover numbers of these two enzymes indicates that at equimolar concentrations, CBS could be the major source of H₂S production, if fully activated by the allosteric regulator AdoMet (Fig. 5, Table 4). At low homocysteine concentrations (10 μM), the value of v/[E] for H₂S generation is predicted to be 0.0249 s⁻¹ and 0.0115 s⁻¹ for CBS and CSE respectively, corresponding to an ~7:3 ratio (CBS:CSE) for H₂S production.
production (Table 4). The contribution of CBS to \( \text{H}_2\text{S} \) production would be ~2-3-fold lower in the absence of allosteric activation by AdoMet, which increases the reaction velocity of the CBS-catalyzed reactions by this factor. Also, in the absence of AdoMet, the \( K_a \) for serine at the PLP site decreases ~4-fold. Thus, the relative contributions of CBS and CSE to \( \text{H}_2\text{S} \) production at low homocysteine concentrations might vary from ~7:3 (full allosteric activation of CBS by AdoMet) to ~2:8 (no activation).

The value of \( v/[E] \) for \( \text{H}_2\text{S} \) generation by CSE is predicted to increase to 0.0217 s\(^{-1}\) and 0.0781 s\(^{-1}\) at 40 \( \mu \)M and 200 \( \mu \)M homocysteine (15) respectively, whereas the comparable values for CBS under conditions of full allosteric activation are largely unaffected (0.0263 s\(^{-1}\) and 0.0269 s\(^{-1}\) at 40 \( \mu \)M and 200 \( \mu \)M homocysteine, respectively). Therefore, under conditions of moderate and severe hyperhomocysteinemia, CSE is predicted to become an increasingly important contributor to the \( \text{H}_2\text{S} \) pool, with the proportion increasing to 45% and 74% respectively at mild and moderate hyperhomocysteinemia. These values represent a lower estimate for the overall contribution of CSE and would be higher if CBS is not fully activated by AdoMet.

Predicted relative contributions of CBS versus CSE to lanthionine generation as a side product in \( \text{H}_2\text{S} \) producing reactions-Lanthionine is a thioether that is produced as a side product in the \( \text{H}_2\text{S} \) generating \( \beta \)-replacement reactions catalyzed by CBS and CSE in which cysteine serves as the substrate (3). At low homocysteine concentrations (10 \( \mu \)M) and equimolar concentrations of both enzymes, the \( v/[E] \) value for reaction 3 is estimated to be 2.9 \( \times \) 10\(^{-4}\) s\(^{-1}\) and 3.9 \( \times \) 10\(^{-4}\) s\(^{-1}\) for CSE and fully AdoMet-activated hCBS, respectively (Fig. 1). Thus, hCBS is predicted to be virtually the sole source (99.99%) of lanthionine formed by condensation of two moles of cysteine. We note that an alternative lanthionine-generating reaction (in which \( \text{H}_2\text{O} \) rather than \( \text{H}_2\text{S} \) is eliminated), namely \( \beta \)-replacement of serine by cysteine might be catalyzed by CSE and CBS. The contribution of this reaction to the total lanthionine pool has not been addressed in this study.

Lanthionine cleavage by CSE-Although lanthionine is a poor substrate for hCBS (\( V_{\text{max}} < 0.05 \) U/mg), human CSE exhibits substantial lanthionine cleavage activity with a \( V_{\text{max}} \) of 1.03 \( \pm \) 0.05 U/mg (\( k_{\text{cat}} = 0.8 \) s\(^{-1}\)) and a \( K_{\text{mapp}} = 0.75 \pm 0.17 \) mM. In comparison, CSE exhibits a \( V_{\text{max}} \) value of 3.1 \( \pm \) 0.1 U/mg (\( k_{\text{cat}} = 2.3 \) s\(^{-1}\)) and a \( K_{\text{m}} \) value of 0.28 \( \pm \) 0.03 mM for cystathionine.

Discussion

Despite the pharmacological interest in modulating \( \text{H}_2\text{S} \) production (28,29) there has been little attention paid to the mechanism of its generation and regulation. In this study, we have investigated the various reactions catalyzed by yeast and human CBS that lead to \( \text{H}_2\text{S} \) production. We have also examined the relative importance of hCBS versus human CSE to \( \text{H}_2\text{S} \) production and the sensitivity of these reactions to the grade of hyperhomocysteinemia.

\( \text{H}_2\text{S} \) production by yCBS-In yeast, ultradian cycling is regulated by chemical signals including \( \text{H}_2\text{S} \) (30) to achieve population synchronization. Interestingly, the \( \text{H}_2\text{S} \) producing sulfate assimilation pathway and respiratory oscillations have been linked to the transsulfuration pathway with intracellular cysteine levels and CSE playing a regulatory role (31).

Yeast CBS catalyzes the production of cystathionine by a \( \beta \)-replacement reaction involving serine and homocysteine (reaction 1). The turnover number of this reaction under \( V_{\text{max}} \) conditions (25.5 s\(^{-1}\)) is ~2-fold lower than for the condensation of cysteine and homocysteine (4, 55.0 s\(^{-1}\)), leading to \( \text{H}_2\text{S} \) production (Table 1). Yeast maintains very low cysteine concentration (0.25-0.45 \( \mu \)M), which is ~1-10% of the intracellular concentration of other amino acids in this organism (4,31). Hence, at intracellular levels of cysteine and serine, reaction 1 is expected to dominate over 4. Of the three \( \text{H}_2\text{S} \) generating reactions catalyzed by yCBS, the \( \beta \)-replacement of cysteine by homocysteine (4) exhibits a higher \( k_{\text{cat}} \) than for reactions 2 and 3 (Table 1). For the bimolecular reactions, the kinetic fits support a ping-pong mechanism, which is consistent with the mechanism previously reported for reaction 1 catalyzed by yCBS (21).
Due to its toxicity to cells at elevated concentrations (32), intracellular cysteine levels are tightly regulated. H$_2$S production by yCBS from cysteine and homocysteine will be sensitive to levels of these amino acids. However, at intracellular concentrations of substrates, the H$_2$S-generating reactions are likely to be competitively inhibited by serine, which is abundant, binding to site 1. The mechanisms by which yCBS is regulated so that it serves either in the transsulfuration pathway or is diverted for H$_2$S production are not known, and merit investigation.

Our studies reveal that yCBS is capable of generating lanthionine (reaction 3), a novel thioether product that is a component of the class of lantibiotics (33). While cyclic lanthionine ketimine has been reported in bovine and human brain (34,35) the presence and fate of this compound in yeast awaits elucidation. Lanthionine like cysteine or cystine, can support the organic sulfur need for growth during the shift from the mycelial to the parasitic yeast form in the dimorphic pathogenic fungus, Histoplasma capsulatum (36). This suggests that lanthionine is transported into yeast cells and metabolized, probably by the action of CSE to cysteine, as discussed below.

$H_2S$ production by hCBS-In contrast to the profligacy of human CSE (15), hCBS has been reported to catalyze H$_2$S formation solely by the $\beta$-replacement of the physiologically relevant substrates cysteine by homocysteine (19). We find that hCBS can generate H$_2$S from cysteine, albeit with lower $k_{cat}$ than from cysteine and homocysteine (Table 1). CBS has also been reported to catalyze H$_2$S generation by $\beta$-replacement of cysteine by several compounds, e.g. $\beta$-mercaptoethanol, dithiothreitol, cysteamine and methanethiol, with concomitant formation of the corresponding thioethers (26).

The canonical reaction catalyzed by hCBS in the transsulfuration pathway is the $\beta$-replacement of serine by homocysteine to generate cystathionine (Fig. 1). The turnover number under substrate saturation ($k_{cat}$) of this reaction (1) is 3.7-fold lower than for cystathionine production from cysteine and homocysteine (4) (Table 1), and could partly be explained by the better leaving group potential of H$_2$S versus H$_2$O. Consequently, at physiological substrate concentrations, the turnover number for reaction 1 is predicted to be 1.3-fold lower than for reaction 4 (Table 2). Thus, reaction 4 represents an alternative route for generating cystathionine at physiologically relevant substrate concentrations, accounting for ~56% of its total synthesis (Fig. 1). However, unlike reaction 1, reaction 4 does not lead to net cysteine synthesis via the transsulfuration pathway since cysteine is consumed to produce cystathionine and then subsequently regenerated by cleavage of cystathionine catalyzed by CSE (Fig. 1). Since the other product of reaction 4 is the signaling molecule H$_2$S, flux through this reaction must be regulated in the cell by mechanisms that await elucidation.

Of the three H$_2$S-producing reactions catalyzed by CBS, reaction 4 is the most efficient, accounting for 96% of the net production (Table 3). In contrast, the $\beta$-elimination (reaction 2) and $\beta$-replacement (3) reactions involving cysteine are of minor importance, representing between 1.6-2.6% of the total H$_2$S derived from hCBS (Table 3).

In contrast to the sensitivity of CSE-catalyzed H$_2$S generation from homocysteine (15), our simulations reveal that H$_2$S production by hCBS is insensitive to the grade of hyperhomocysteinemia (Table 2). Increasing homocysteine from normal to moderate to severe elevated levels has virtually no effect on the $k_{cat}$ for reaction 4 for H$_2$S production (Table 2). The insensitivity to homocysteine concentrations results from the relatively low $K_d$ for serine binding to site 1 ($K_d$~56 µM) compared to the physiological concentration of serine (560 µM). Thus, serine competes effectively with cysteine, which is present at lower concentrations and exhibits a higher $K_d$, and thus limits the three H$_2$S generating reactions since they involve binding of cysteine to site 1 (Fig. 6). Unlike CSE, homocysteine appears to be unable to bind to site 1 in hCBS and the enzyme does not catalyze reactions at the $\gamma$-carbon. Consequently, the only reaction rate that is boosted by an increase in homocysteine concentration is the $\beta$-replacement of serine by homocysteine (reaction 1) whose turnover number is predicted to increase 3.8- and 14.7-fold under moderate and severe
hyperhomocysteinemia respectively (Table 2). This enhancement would only be relevant in hyperhomocysteinemia conditions resulting from a deficiency in enzymes other than hCBS.

The insensitivity of H₂S formation by CBS to homocysteine suggests that CSE will be primarily responsible for enhanced H₂S production under conditions of hyperhomocysteinemia (Table 4, Fig.5). We have estimated the relative contribution of CBS and CSE to the net H₂S pool, assuming equivalent amounts of both enzymes and full activation of CBS by AdoMet. Thus, under normal conditions (10 µM homocysteine), CSE accounts for ~32% of the H₂S produced by the transsulfuration pathway and this increases to ~45 and ~74% under moderate and severe hyperhomocysteinemia conditions. However, in tissues where one or the other enzyme is absent or present at greatly reduced levels, their relative contributions to H₂S biogenesis will clearly be different. Furthermore, depending on the extent of CBS activation by AdoMet, its relative contribution to the H₂S pool could vary between ~25-70% in cells containing equimolar CSE.

Mutations in CBS are the most common cause of severe hyperhomocysteinemia in comparison to defects elsewhere in the metabolic pathway (37). In homocystinurics with CBS deficiency, CSE will not only be the primary source of H₂S, but is predicted to generate higher H₂S levels due to increased accumulation of homocysteine (15). Under conditions of elevated homocysteine due to deficiencies elsewhere in the methionine cycle, i.e. not in CBS, or higher flux through the transsulfuration pathway (e.g. after a methionine-rich meal), cystathionine production is predicted to increase significantly (Fig. 1, Table 2). This could lead in turn to enhanced production of cysteine and stimulate glutathione production, since cysteine is the limiting substrate for the synthesis of this antioxidant. The activity of γ-glutamylcysteine synthetase, the committing step in the synthesis of glutathione, is regulated by feedback inhibition by glutathione (38). Interestingly, H₂S has been reported to enhance the activity of γ-glutamylcysteine synthetase (39).

Rat CBS has been reported to generate lanthionine from a mixture of cysteine and serine (25). Here, we show that lanthionine (reaction 3) can also be produced by the condensation of two moles of cysteine (Table 1). The estimated turnover number for lanthionine production by hCBS is ~13,000-fold higher than by human CSE. In contrast, CSE catalyzes the cleavage of lanthionine to cysteine and pyruvate more efficiently (k_cat~0.8 s⁻¹) than CBS (k_cat<0.05 s⁻¹). Thus, lanthionine concentrations are expected to be higher in tissues in which CSE levels and/or activity are low. Our results demonstrate that lanthionine derived from cysteine is produced almost exclusively (99.99%) by CBS, thus linking the origin of this metabolite to cysteine metabolism and CBS.

**Implications of relaxed substrate specificity in the transsulfuration pathway**—The profligacy of CBS and CSE lead to a much greater complexity of transsulfuration reactions catalyzed by these enzymes than previously appreciated and expand the sulfur metabolome. The relative insensitivity of CBS to the grade of hyperhomocysteinemia contrasts with that of CSE. These results imply that CSE plays a heretofore unrecognized role in intracellular homocysteine management both in the peripheral vasculature where CBS levels are low or the enzyme is absent, and under hyperhomocysteinemic conditions. Evidence for the role of CSE in homocysteine homeostasis is provided by the ~18-fold increase in homocysteine levels in aorta and heart in CSE⁻/⁻ mice (1). The metabolic functions of lanthionine, which is predicted to be enriched in tissues with low CSE activity, and of homolanthionine, await further elucidation. The capacity of the transsulfuration pathway to catalyze a multitude of reactions raises important questions about how these reactions are regulated to meet cellular needs for the varied products.
Acknowledgements
The authors thank Dr. Weidong Zhu (University of Nebraska-Lincoln) for assistance with performing the mass spectrometric analysis of our samples.
REFERENCES


Table 1
Kinetic parameters determined for reactions catalyzed by yeast or human CBS.

<table>
<thead>
<tr>
<th>Reaction</th>
<th>#</th>
<th>V_{max}, U^{b}/mg</th>
<th>K_{m1}, mM</th>
<th>K_{i}, mM</th>
<th>n</th>
<th>k_{cat}, s^{-1}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cystathionine generation from Ser + Hcys</td>
<td>1</td>
<td>24.3 ± 2.4</td>
<td>3.4 ± 0.2(^{d})</td>
<td>-</td>
<td>-</td>
<td>2.5 ± 2.5</td>
</tr>
<tr>
<td>H₂S generation from Cys</td>
<td>2</td>
<td>3.7 ± 1.1</td>
<td>3.6 ± 1.7</td>
<td>33 ± 3.7</td>
<td>-</td>
<td>3.8 ± 1.0</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>9.0 ± 1.3</td>
<td>33 ± 3.7(^{d})</td>
<td>2.4 ± 0.5</td>
<td>8.1 ± 1.2</td>
<td></td>
</tr>
<tr>
<td>H₂S generation from Cys + Hcys</td>
<td>4</td>
<td>60 ± 2.4</td>
<td>3.6 ± 1.7(^{e})</td>
<td>-</td>
<td>0.13 ± 0.02(^{d})</td>
<td>0.9 ± 0.1 55 ± 2.2</td>
</tr>
</tbody>
</table>

Yeast CBS

<table>
<thead>
<tr>
<th>reaction</th>
<th>#</th>
<th>V_{max}, U^{b}/mg</th>
<th>K_{m1}, mM</th>
<th>K_{i}, mM</th>
<th>n</th>
<th>k_{cat}, s^{-1}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cystathionine generation from Ser + Hcys</td>
<td>1</td>
<td>5.05 ± 0.3</td>
<td>2.76 ± 0.5(^{d})</td>
<td>-</td>
<td>-</td>
<td>5.3 ± 0.3</td>
</tr>
<tr>
<td>Serine generation from Cys</td>
<td>2</td>
<td>0.34 ± 0.04</td>
<td>6.8 ± 1.7</td>
<td>42 ± 10.5</td>
<td>-</td>
<td>0.96 ± 0.04</td>
</tr>
<tr>
<td>Lanthionine generation from Cys</td>
<td>3</td>
<td>0.77 ± 0.07</td>
<td>6.8 ± 1.7(^{e})</td>
<td>45.6 ± 5.6(^{d})</td>
<td>2.1 ± 0.4</td>
<td>0.86 ± 0.07</td>
</tr>
<tr>
<td>H₂S generation from Cys</td>
<td>2</td>
<td>0.46 ± 0.08</td>
<td>6.8 ± 1.7(^{e})</td>
<td>27 ± 3.7</td>
<td>0.48 ± 0.08</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>0.82 ± 0.08</td>
<td>27.3 ± 3.7(^{d})</td>
<td>1.3 ± 0.3</td>
<td>0.86 ± 0.08</td>
<td></td>
</tr>
<tr>
<td>H₂S generation from Cys + Hcys</td>
<td>4</td>
<td>18.7 ± 2.6</td>
<td>6.8 ± 1.7(^{e})</td>
<td>-</td>
<td>3.2 ± 1.3(^{d})</td>
<td>0.74 ± 0.1 19.6 ± 2.7</td>
</tr>
</tbody>
</table>

Human CBS

\(^{a}\)All values reported in the table were obtained from the fit of a plot containing data from at least three independent experiments and are represented ±S.E.. \(^{b}\)One unit of activity is defined as one micromole of product generated min\(^{-1}\) at 37°C. \(^{c}\)The activity was determined in the radiolabeled assay. \(^{d}\)The K_{m} value corresponds to K_{m2}, i.e. for substrate binding to site 2. \(^{e}\)The value of K_{m1} was fixed to determine the kinetic parameters V_{max2}, K_{m2} and n.
**Table 2**

Kinetic parameters for the hCBS-catalyzed reactions at varying homocysteine concentrations estimated by simulations\(^a\)

<table>
<thead>
<tr>
<th>Substrates</th>
<th>#</th>
<th>( V_{\text{max}} ) (^b) U/mg</th>
<th>( K_{m1}, K_{m2} ) (^c) (mM)</th>
<th>( v/[E] ) (^d) (sec(^{-1}))</th>
<th>( v/[E] ) (^d) (sec(^{-1}))</th>
<th>Fold-change(^e)</th>
<th>( v/[E] ) (^d) (sec(^{-1}))</th>
<th>Fold-change(^e)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ser + Hcys</td>
<td>1</td>
<td>5.1 ± 0.3</td>
<td>2.0±0.3, 2.8±0.5</td>
<td>185×10(^{-4})</td>
<td>700×10(^{-4})</td>
<td>3.8</td>
<td>2747×10(^{-4})</td>
<td>14.7</td>
</tr>
<tr>
<td>Cys</td>
<td>2</td>
<td>0.46 ± 0.08</td>
<td>6.8±1.7</td>
<td>6.4×10(^{-4})</td>
<td>6.4×10(^{-4})</td>
<td>0.99</td>
<td>6.8×10(^{-4})</td>
<td>0.96</td>
</tr>
<tr>
<td>Cys+Cys</td>
<td>3</td>
<td>0.82 ± 0.08</td>
<td>6.8±1.7, 27.3± 3.7</td>
<td>3.9×10(^{-4})</td>
<td>3.9×10(^{-4})</td>
<td>0.99</td>
<td>3.9×10(^{-4})</td>
<td>0.97</td>
</tr>
<tr>
<td>Cys + Hcys</td>
<td>4</td>
<td>18.7 ± 2.6</td>
<td>6.8± 1.7, 3.2±1.3</td>
<td>239×10(^{-4})</td>
<td>253×10(^{-4})</td>
<td>1.06</td>
<td>253×10(^{-4})</td>
<td>1.09</td>
</tr>
</tbody>
</table>

\(^a\)The reaction numbers correspond to those shown in Fig. 2. \(^b\)One unit corresponds to 1 µmole product formed min\(^{-1}\). The \( K_m \) and \( V_{\text{max}} \) values were determined as described under Experimental Procedures and reported in Table 1. \(^c\)In reactions involving two substrates, the order of the \( K_m \) values reflects the substrate order in the first column. \(^d\)The values for the turnover numbers at varying concentrations of homocysteine and physiological concentrations of serine (560 µM) and cysteine (100 µM) were obtained as described under Experimental Procedures considering a ping-pong mechanism for the bimolecular reaction and the Hill coefficients (n) reported in Table 1. \(^e\)Fold change refers to the change in \( v/[E] \) with respect to normal conditions, i.e. 10 µM homocysteine, which is assigned a value of 1 for each reaction.
Table 3

The relative contributions of the H₂S generating reactions at varying concentrations of homocysteine as predicted by kinetic simulations

<table>
<thead>
<tr>
<th>Substrate</th>
<th>Reaction #</th>
<th>10 µM Hcys</th>
<th>40 µM Hcys</th>
<th>200 µM Hcys</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cys</td>
<td>2</td>
<td>2.6</td>
<td>2.4</td>
<td>2.3</td>
</tr>
<tr>
<td>Cys + Cys</td>
<td>3</td>
<td>1.6</td>
<td>1.5</td>
<td>1.4</td>
</tr>
<tr>
<td>Cys + Hcys</td>
<td>4</td>
<td>95.8</td>
<td>96.1</td>
<td>96.3</td>
</tr>
</tbody>
</table>

*The reaction numbers correspond to those shown in Figure 2. “%” refers to the percent contribution of each reaction to net H₂S production at each concentration of homocysteine in the presence of 560 µM serine and 100 µM cysteine.*
Table 4

The relative contributions of CBS and CSE to net H\textsubscript{2}S production via the transsulfuration pathway at varying homocysteine concentrations predicted by kinetic simulations\textsuperscript{a}

<table>
<thead>
<tr>
<th>enzyme</th>
<th>substrate</th>
<th>10 (\mu\text{M}) HCys</th>
<th>40 (\mu\text{M}) HCys</th>
<th>200 (\mu\text{M}) HCys</th>
</tr>
</thead>
<tbody>
<tr>
<td>CBS</td>
<td>Cys</td>
<td>2.8 % 68 % total</td>
<td>2.1 % 55 % total</td>
<td>0.94 % 26 % total</td>
</tr>
<tr>
<td>CBS</td>
<td>Cys+HCys</td>
<td>65.6 %</td>
<td>52.6 %</td>
<td>24.6 %</td>
</tr>
<tr>
<td>CSE</td>
<td>HCys</td>
<td>9.3 % 32 % total</td>
<td>28.5 % 45 % total</td>
<td>67.1 % 74 % total</td>
</tr>
<tr>
<td>CSE</td>
<td>Cys+HCys</td>
<td>0.005 %</td>
<td>0.03 %</td>
<td>0.19 %</td>
</tr>
</tbody>
</table>

\textsuperscript{a}The % contributions for CBS and CSE were calculated using the turnover numbers reported in Table 2 for CBS and the previously reported values for CSE (15) as described under Experimental Procedures.
FIGURE LEGENDS

FIGURE 1. The diversity of reactions catalyzed by the transulfuration pathway. The turnover numbers ($v/[E]$) estimated at physiological substrate concentrations, i.e. 10 µM homocysteine, 100 µM cysteine, 560 µM serine and 5 µM cystathionine, are shown in parentheses for each reaction. The thick arrows highlight reactions that are sensitive to elevated levels of homocysteine. The fold change represents the fold-increase in the turnover number of a given reaction under conditions of severe hyperhomocysteinemia (200 µM homocysteine).

FIGURE 2. Reactions catalyzed by CBS leading to cystathionine, lanthionine and H$_2$S-generation that were characterized in this study. The dotted arrows in reactions 2 and 5 denote reactions for which our study did not find evidence.

FIGURE 3. Kinetics of H$_2$S generation by yCBS. (A) Kinetics of H$_2$S generation from cysteine and the contributions of reactions 2 and 3 catalyzed by yCBS. The kinetic data for H$_2$S generation from cysteine (reactions 2+3) are shown by open circles. Each data point represents the mean (± S.D.) of at least three independent experiments. The data were analyzed as described under experimental procedures and the kinetic parameters obtained from these plots are shown in Table 1. The contributions of the component reactions ($v_2$ and $v_3$) to the net rate of H$_2$S generation are shown by gray lines. (B) Kinetics of H$_2$S production (○) observed at 15 mM cysteine and varying concentrations of homocysteine. Each data point represents the mean (± S.D) of three independent experiments. The relative contributions of the individual reactions ($v_2$, $v_3$ and $v_4$) to net rate of H$_2$S production were simulated using the kinetic parameters reported in Table 1 and as described under Experimental Procedures.

FIGURE 4. Kinetics of H$_2$S generation by hCBS. (A) Dependence of H$_2$S generation (reactions 2+3) on cysteine concentration. The experimental data are shown by open circles while the lines represent the fits obtained as described under Experimental Procedures for the contributing reaction. (B) Kinetic dependence of serine (reaction 2, △) and lanthionine (reaction 3, ○) generation on cysteine concentration. The lines represent fits obtained as described under Experimental Procedures. (C) Kinetics
of H₂S production (○) observed at 20 mM cysteine and varying concentrations of homocysteine. Each data point represents the mean (± S.D) of three independent experiments. The relative contributions of the individual reactions (ν₂, ν₃ and ν₄) to the net rate of H₂S production were simulated using the kinetic parameters reported in Table 1 and as described under Experimental Procedures.

FIGURE 5. Relative contributions of fully activated hCBS and CSE to H₂S generated via the transsulfuration pathway determined by simulations (A) The relative proportions of fully activated CBS- and CSE-derived H₂S at three concentrations of homocysteine and physiological concentrations of serine, cysteine and cystathionine. (B) The relative contributions of fully activated CBS and CSE to net H₂S production via the transsulfuration pathway at three concentrations of homocysteine mimicking normal (10 µM), moderate (40 µM) and severe (200 µM) hyperhomocysteinemia.

FIGURE 6. Schematic representation for the sensitivity of the CSE- but not CBS-catalyzed H₂S production to hyperhomocysteinemia. Unlike CSE, the PLP-binding pocket of CBS binds serine or cysteine but not homocysteine.
FIGURE 1

Serine + Homocysteine

Cysteine

H₂S

CBS

H₂S

Cysteine + Homocysteine

Serine

(0.64 × 10⁻³ s⁻¹)

Lanthionine

(0.39 × 10⁻³ s⁻¹)

Cystathionine

(18.5 × 10⁻³ s⁻¹)

× 14.7 ↑

Cysteine

H₂S

CSE

H₂S

Cysteine + Homocysteine

Homocysteine

Pyruvate

(8.1 × 10⁻³ s⁻¹)

Lanthionine

(0.029 × 10⁻⁴ s⁻¹)

Cysteine

(39 × 10⁻³ s⁻¹)

α-ketobutyrate

(3.4 × 10⁻³ s⁻¹)

Homolanthionine

(42 × 10⁻⁴ s⁻¹)

Cystathionine

(1.8 × 10⁻⁴ s⁻¹)

× 18 ↑

× 234 ↑

× 114 ↑
FIGURE 2

Reaction (1) β-replacement

Reaction (2) α,β-elimination

Reaction (3) β-replacement

Reaction (4) β-replacement

Reaction (5) α,γ-elimination
FIGURE 3

A

B

[Image of a graph showing Michaelis-Menten kinetics for enzyme activity with [L-Cys] mM on the x-axis and V H2S U/mg on the y-axis.]

[Image of a graph showing the inhibition of enzyme activity with [L-Hcys] mM on the x-axis and V H2S U/mg on the y-axis with lines representing different concentrations of inhibitor.]
FIGURE 4

A

B

C

\[ \text{v}_{\text{HgS}, \text{ U/mg}} \]

\[ \text{v}_{\text{Pr-glu}} \text{, U/mg} \]

\[ \text{v}_{\text{HgS}, \text{ U/mg}} \]

\[ \text{[L-Cys], mM} \]

\[ \text{[L-Cys], mM} \]

\[ \text{[L-Hcys], mM} \]
FIGURE 5

A

B

contribution to H₂S production, %

[HCys], µM

contribution to H₂S production, %

[HCys], µM
FIGURE 6

CBS

PLP
serine
PLP
cysteine
PLP
homocysteine

CSE

PLP
cystathionine
PLP
cysteine
PLP
homocysteine